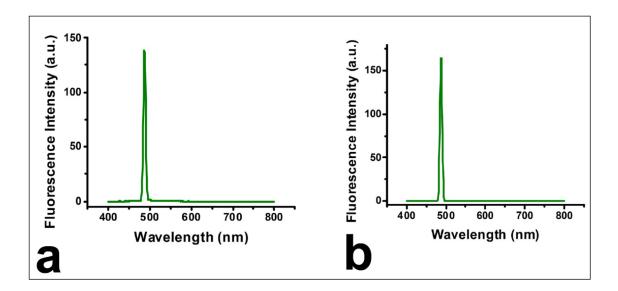
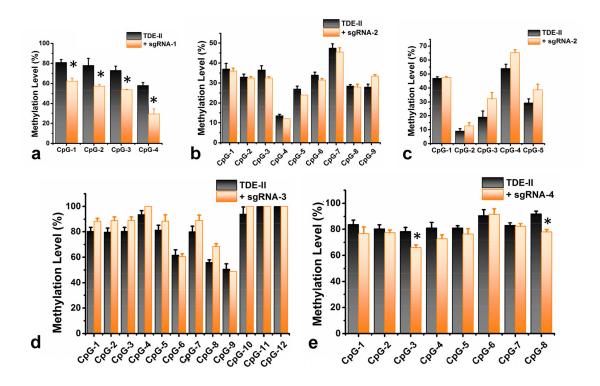
CRISPR-dCas9 mediated TET1 targeting for selective DNA demethylation at *BRCA1* promoter

SUPPLEMENTARY DATA

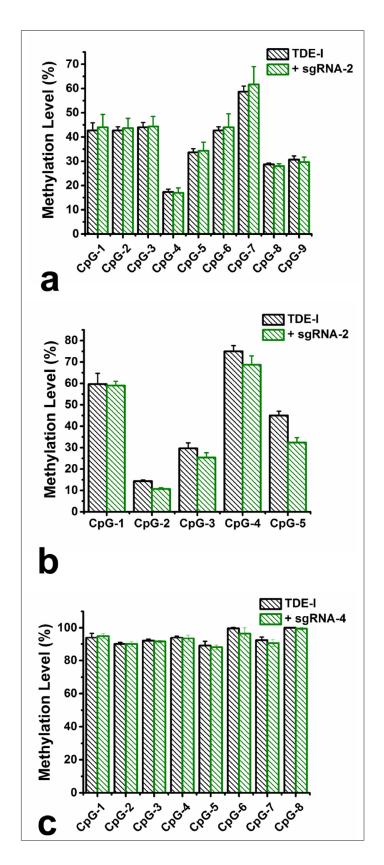
See Supplementary Sequence File: 1



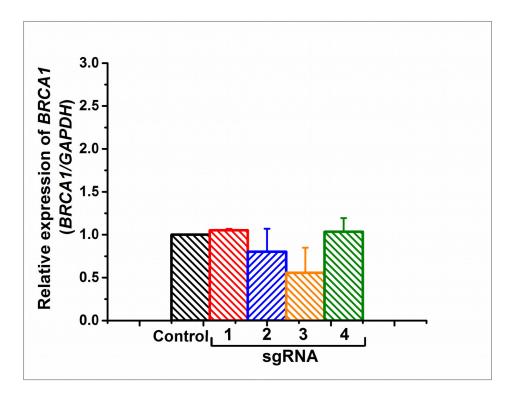
Supplementary Figure S1: The total protein was extracted from the co-transfected (combination of TET1-dCas9 plasmids and sgRNAs) cells, and the emission spectra of the fluorescent tags (EGFP) in the fusion proteins were recorded using a fluorimeter. A slight shift in EGFP emission also suggests the possible formation of TET1-dCas9-EGFP fusion proteins namely TDE-I and TDE-II. Representative emission spectra is presented for TDE-I in HeLa a. and MCF7 b. cell free extracts.



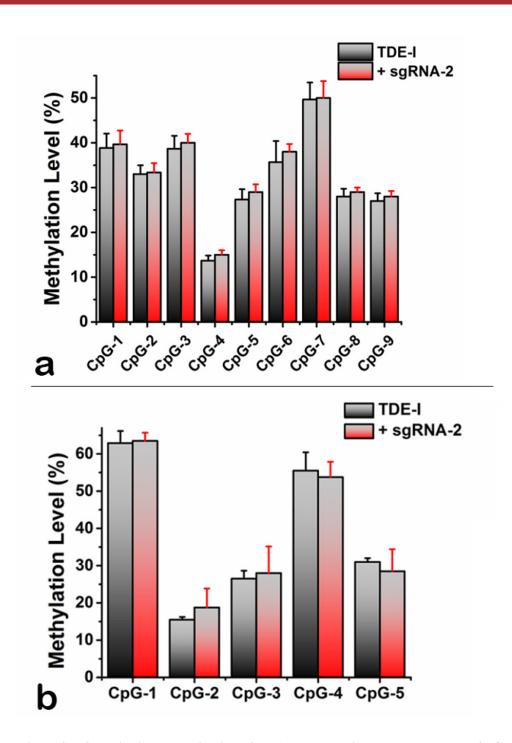
Supplementary Figure S2: Quantitative determination of DNA demethylation levels at the target CpG sites in BRCA1 promoter. The levels of DNA methylation were determined by pyrosequencing in HeLa cells, treated with TDE-II and different combinations of sgRNAs (in orange), compared to only TDE-II treated ones (in black) (a-e). A p value of < 0.05 was considered statistically significant for all the obtained data.



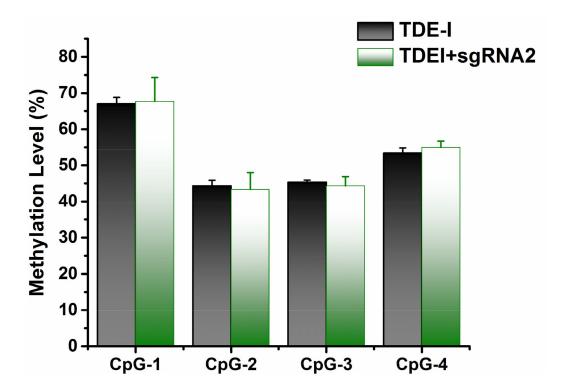
Supplementary Figure S3: Quantitative determination of DNA demethylation level at the target CpG sites in *BRCA1* **promoter.** The levels of DNA methylation were determined by pyrosequencing in MCF7 cells, treated with the combination of TDE-I and sgRNA-2 (a, b), and the combination of TDE-I with sgRNA-4 in MCF-7 cells.



Supplementary Figure S4: The gene expression of BRCA1 was determined with qPCR analysis followed by the cotreatment with TDE-II and different combinations of sg-RNAs in HeLa cells.



Supplementary Figure S5: Quantitative determination of DNA demethylation level at the target CpG sites in *BRCA1* **promoter.** The levels of DNA methylation were determined by pyrosequencing in HeLa cells, treated with the combination of inactive TET1 fusion protein and sgRNA-2 (a, b).



Supplementary Figure S6: Quantitative determination of DNA demethylation levels at the target CpG sites at global LINE-1 repeat sequence element. The levels of DNA methylation were determined by pyrosequencing in HeLa cells, treated with TDE-I and compared to the combination of TDEI + sgRNA-2 (a-e). A p value of < 0.05 was considered statistically significant for all of the obtained data

Supplementary Table S1: Primers used in this study for PCR amplification and sequencing analysis of the fusion-protein construct (TET1CD- dCas9-EGFP), all primers listed are 5' to 3'

TET1CD- dCas9-EGFP (TDE) fusion protein construct				
PCR primers				
Insert name		Forward	Reverse	
TDE-I	TET1CD	atcgttGGTACCGCCACCATGGAACT GCCCACCTGCAGCTGTCTTGAT	atcgttGGATCCGAC CCAATGGTTAT AGGGCCCCGCAACGTG	
	dCas9	ATCGTTGGATCC AGCGGAAGTACACCCGCA ATGGCTAGCCCCAAA AAGAAGAGGAAAGTG	atcctaTCTAGA ACCTACCTTGC GCTTTTTCTTGGGAG	
TDE-II	TET1CD	ategttGGTACC GCCACC ATGGAACTGCC CACCTGCAGCTGTCTTGAT	atcgttGGATCC GACCCAATGGTT ATAGGGCCCCGCAACGTG	
	dCas9	ategttGGATCC GCCACC ATGGCTAGCCCC AAAAAGAAGAGGAAAGTG	atcctaTCTAGA ACCTACCTTG CGCTTTTTCTTGGGAG	
		Sequencing primers		
Primer name	Sequence (5' to 3')	Primer name	Sequence (5' to 3')	
TDE_SP1	GAGGGGTTTTATGCGATG	TDE_SP6	CGTGAACACTGAA ATCACCAAG	
TDE_SP2	TCCAAGCTCTCCCTTACATGA	TDE _SP7	GTACAACGAGCT GACCAAGGTG	
TDE_SP3	ACCTTAGGGAGT AACACTGAGACC	TDE _SP8 GTATTCTCCAGACCG TGAAAGTC		
TDE_SP4	ACATTGATGAGTAT TGGTCAGACAG	TDE _SP9	CGCGAGGTG AAGGTGATTA	
TDE_SP5	ATCCCATCTTCGGTAATATCGT	TDE _SP10	GAACTGCTGGGT ATCACCATTAT	
Sequence (3	' to 5')			
TDE_SP11		TGTACAGCTCGTCCATGC		

The restriction sites are represented in green, linkers in red, and the sites complementary to the inserts in black font.

Supplementary Table S2: Primers used in this study for PCR amplification and sequencing analysis of the fusion-protein dCas9- TET1CD (inactive)-EGFP, all primers listed are 5' to 3'

	dCas9- TET1CD (inactive)- EGFP fusion protein construct			
PCR primers				
	Insert name	Forward	Reverse	
	dCas9	atcgttCCATGGCTAGCCCC AAAAAGAAGAGGAAAGTG	gctagcGGCGCGCCCACCTA CCTTGCGCTTTTTCTTGGGAG	
dCas9-TET1CD (inactive)-EGFP	TET1CD (inactive)	atcgttGCTAGC AGCGGA AGTACACCCGCA ATGGAACTGCCCA CCTGCAGCTGTCTTGAT	atcctaCCTGCAGGGACCCAAT GGTTATAGGGCCCCGCAACGTG	
	EGFP	atcctaCCTGCAGG AAGCGGAAGTACACCCGCA ATGGTGAGCAAGGGCGAGG	tctacaaaGCGGCCGCCTACT TGTACAGCTCGTCCATG	

The restriction sites are represented in green, linkers in red, and the sites complementary to the inserts in black font.

Supplementary Table S3: Primers used in this study for the PCR amplification of *BRCA1* after bisulfite conversion of the HeLa genomic DNA. The target region was amplified in two fragments with 2 sets of primers.

BSP-PCR primers				
	Forward	Reverse	Product size (bp)	Annealing Temp (Tm)
Fragment-1 (F1)	/ 5BiosG /GGGTTAGT TAGGGGTGGGGTTA	CCTCTCCCTCCACACTTC	380	58 °C
Fragment-2 (F2)	TATTTTGGTAGTG TTGGAGGAGTT	/ 5BiosG /AACCACCCT AAAACTCACAAAAATTAAA	400	56 °C
	Pyrosequencing primers	Target sequencing CpG sites (h	ighlighted in y	vellow)
F1-S1	*CTCTCATCCTATCACTAAAA	CGATTTTTTCGTG TTTTCGGATAGTTAATCG		
F1-S2	*CAATAACCAACT AAAAAACTCCTC	CGAGGTGATAACGTGTTAGT AGTTTTCGTTCGTTTTTCGGC TTTCGGTTTTTGGCGTTTATTT	GTTT	
F2-S1	GTGTTGGAGGAGTTT	CGTTATTGCGTTGTGGGGGT TTTTGGGTTGGTCGAAGTTA TCGGTTTTTTTTGTTTGCG		
F2-S2	TGTGGGAATTGGGGT	CGCGTAGCGTTCGTTAGTTA TTAGGTGGGCGCGGGTTTAC TTCGTATTTTCGGTTTCG		
F2-S3	GTTTAGGGTAGTTAGGGG	CGGGTTAGTAGTTGTAGAGC TGCGTCGGGTTTTTTAGTAT TCGGTTCGTTTGTATTTCGTT ATTTTTATCGGGTTTTAGTCC	ΓG ΓTGA	

Pyrosequencing was then carried out to quantify the methylation percentage at each CpG sites of the analyzed regions. All the primers listed are 5' to 3'. Bisulfite-PCR (BSP) primers were 5'-biotinylated in either forward or reverse primer.

^{*}Sequencing primer binds to the reverse strand. The sequence of corresponding CpG sites of these sequencing primers hence can be read in the reverse direction of the actual sequence.

Supplementary Table S4: Primers used to study transcript quantification of *BRCA1* relative to the endogenous control GAPDH by qPCR

	Forward	Reverse
BRCA1	CAAGGAACCAGGGATGAAATCAG	ATGGCTCCACATGCAAGTTTG
GAPDH	CAGCCTCAAGATCATCAGCA	TGTGGTCATGAGTCCTTCCA

All the primers listed are 5' to 3'.

Supplementary Table S5: Primers used for qPCR to determine locus specific 5-hydroxymethylation (5-hmC) at the target site of *BRCA1* promoter

Treated with TDE-I plus sgRNA-2			
Primer sequence Covered 5-hmC sites [†] (Amplicon size: 270 bp)			
Forward	GAGCCCTTCGTGTTCTGAGG	GCGCCGGAGAGTTGGAGAGTCTGTGGT TCAGAATGCGAGGTGACAACGTGCTAGC AGCCCTCGCTCGCTCTCGGCGCCTCCTCGGCCTT GGCGTCCATTCTGGCCGTGCTGGAGGA GCCCTTCAGCCCGCCACTGCGC	
Reverse	GGTGTGGGAACTGGGGCT		

All the primers listed are 5' to 3'.